

SALVIFARIN, X-RAY STRUCTURE DETERMINATION OF A *CIS* NEO-CLERODANE DITERPENOID FROM *SALVIA FARINACEA*

LILIANA EGUREN, JOSÉ FAYOS, AUREA PERALES, GIUSEPPE SAVONA* and BENJAMÍN RODRÍGUEZ†

Departamento de Rayos X, Instituto 'Rocafolano', CSIC., Serrano 119, Madrid-6, Spain; *Istituto di Chimica Organica dell'Università, Archirafi 20, 90123 Palermo, Italy; †Instituto de Química Orgánica, CSIC., Juan de la Cierva 3, Madrid-6, Spain

(Received 16 June 1983)

Key Word Index—*Salvia farinacea*; Labiatae; diterpenoid; *cis* neo-clerodane; salvifarin; revised structure.

Abstract—The structure and absolute configuration of salvifarin, a neo-clerodane diterpenoid isolated from *Salvia farinacea*, have been established by X-ray diffraction analysis. This result modifies the structure previously assigned to this compound.

In a previous communication [1] we reported the isolation and structure determination of two new diterpenoids, salvifarin and salvifarin, from *Salvia farinacea* Benth. (Labiatae). On the basis of extensive ^1H and ^{13}C NMR spectroscopic studies we assigned structure 1 for salvifarin, but its configuration at the C-12 centre was suggested as 12*R* only on biogenetic grounds. This prompted us to obtain the X-ray diffraction molecular structure of salvifarin to definitely establish this point. Figure 1 shows the X-ray absolute molecular structure of the diterpenoid, in which the configuration at C-12 is *R*, as it was put forward earlier [1], but the configurations of the C-10 hydrogen atom and of the C-1, C-2-oxirane ring are opposite to those previously reported [1]. Thus, salvifarin possesses the structure of *cis* neo-clerodane depicted in formula 2. Its ring A is roughly between a half-chair and twist conformation, whereas ring B has a chair distorted to envelope conformation. The conformational parameters of salvifarin (2) have been calculated in the same way as those reported [2] for some neo-clerodane diterpenoids, and these values are: $\theta_A = 114^\circ$, $\phi_A = 91^\circ$, $Q_A = 0.32 \text{ \AA}$; $\theta_B = 28^\circ$, $\phi_B = 303^\circ$ and $Q_B = 0.67 \text{ \AA}$.

It is important to note that although salvifarin (2) is a *cis* neo-clerodane diterpenoid, and its C-19 methylene and C-20 acetal hydrogen atoms are more distant than in a *trans* neo-clerodane structure, there exists a strong NOE between these protons [1].

EXPERIMENTAL

For isolation of salvifarin (2) from *Salvia farinacea* and its spectroscopic data, see ref. [1].

X-ray structure determination of salvifarin (2). Crystals of salvifarin ($\text{C}_{20}\text{H}_{20}\text{O}_6$) are orthorhombic, $P2_12_12_1$, $Z = 4$, with $a = 20.988$ (2), $b = 9.280$ (1) and $c = 8.6088$ (4) Å, $D_c = 1.412 \text{ g/cm}^3$ and $\mu = 8.24 \text{ cm}^{-1}$. Graphite-monochromatic $\text{CuK}\alpha$ radiation ($\lambda = 1.5418 \text{ \AA}$) was selected to measure the intensity of the 1733 independent Friedel pairs up to $\theta = 67^\circ$, on a crystal of $0.16 \times 0.22 \times 0.24 \text{ mm}$. With the aid of an automatic diffractometer, each reflexion was scanned 1.2° in 0.5 min, by the $\omega/2\theta$ mode, and individual background was also measured. No intensity decay was observed during the experiment. The crystal

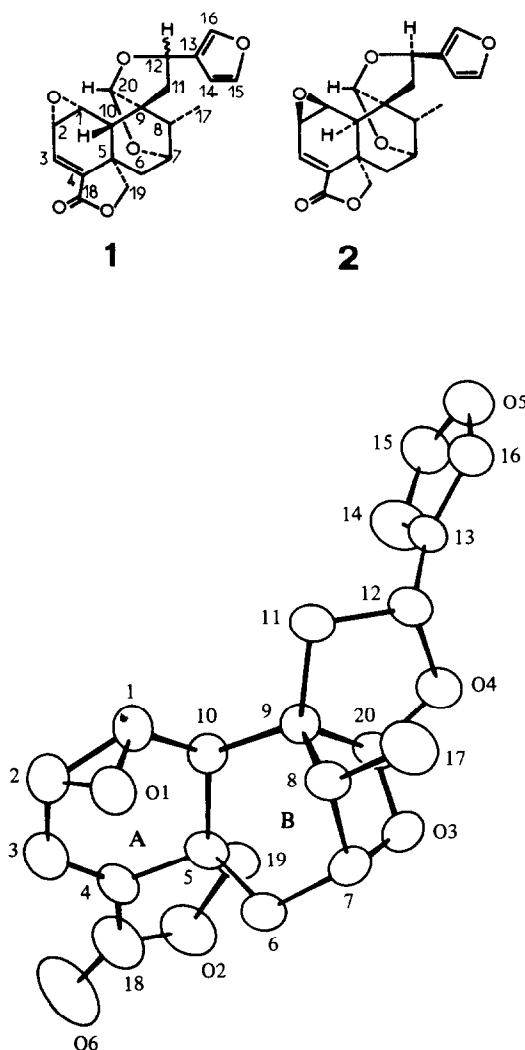


Fig. 1. X-ray molecular model of salvifarin (2).

structure was solved by MULTAN [3], with the 254 E's > 1.42 . After refinement with the 1555 $2\sigma(I)$ observed intensities and location of the hydrogen atoms on a difference map, a weighting scheme was selected to have no dependence of $\langle w\Delta^2 F \rangle$ vs. $\langle F_o \rangle$ and vs. $\langle \sin\theta/\lambda \rangle$. A weighted full matrix L.S. anisotropic refinement (fixed isotropic for H atoms) using the 1555 observed Friedel pairs converged to $R = 4.5\%$ and $R_w = 5.7\%$ [4].

The neo-clerodane absolute configuration of salvifarin (Fig. 1 and 2) was determined comparing the 84 Bijvoet pairs with $\Delta F_c > 0.08$ and with less experimental error, that is $F_o > 10\sigma(F_o)$, $4 < F_o < 25$ and $0.22 < \sin\theta/\lambda < 0.58$. The averaged Bijvoet difference was 0.359 for the right enantiomer vs. 0.434 for the wrong one*.

*A list of atomic parameters, bond distances and angles, torsion angles and $F_o - F_c$ tables are deposited at the Cambridge Crystallographic Data Centre.

Acknowledgement—We thank Professor S. García-Blanco for his support and Centro de Cálculo del Ministerio de Educación y Ciencia, Spain, for computer facilities. The financial support of the Spanish Foreign Ministry for travel facilities between Italy and Spain is gratefully acknowledged.

REFERENCES

1. Savona, G., Raffa, D., Bruno, M. and Rodríguez, B. (1983) *Phytochemistry* **22**, 784.
2. Eguren, L., Perales, A., Fayos, J., Rodríguez, B., Savona, G. and Piozzi, F. (1982) *J. Org. Chem.* **47**, 4157.
3. Main, P. (1980) *MULTAN-80*. Department of Physics, University of York, U.K.
4. Stewart, J. M., Kundell, F. A. and Balwin, J. C. (1970) *The X-Ray 70 System*. Computer Science Center, University of Maryland, U.S.A.

Phytochemistry, Vol. 23, No. 2, pp. 467–468, 1984.
Printed in Great Britain.

0031-9422/84 \$3.00 + 0.00
© 1984 Pergamon Press Ltd.

A COUMARIN ACETYLGLUCOSIDE FROM *VIBURNUM SUSPENSUM*

TETSUO IWAGAWA and TSUNAO HASE

Department of Chemistry, Faculty of Science, Kagoshima University, Korimoto 1-21-35, Kagoshima 890, Japan

(Received 9 August 1983)

Key Word Index—*Viburnum suspensum*; Caprifoliaceae; coumarin; 2',6'-O-diacetylscopolin.

Abstract—A new acetylated coumarin glucoside has been isolated from the leaves of *Viburnum suspensum* and determined as 2',6'-O-diacetylscopolin on the basis of spectral and chemical evidence.

INTRODUCTION

Recently, we have isolated two known flavonol glycosides [1] and several bitter iridoids [2] from the leaves of *Viburnum suspensum* L. Further chromatographic examination of the leaves of the plant gave a new acetyl-coumarin glucoside (1).

RESULTS

Compound **1** was crystallized as needles, mp 178–179.5°, $[\alpha]_D^{25} - 100^\circ$ from methanol. The molecular formula $C_{20}H_{22}O_{11} \cdot 1/2 H_2O$ was determined on the basis of the elementary analysis and mass spectrum. Absorption bands at 1740, 1620 and 1570 cm^{-1} in the IR spectrum and absorption maxima at 227 nm ($\epsilon 10800$), 286 nm ($\epsilon 5600$) and 328 nm ($\epsilon 7600$) in the UV spectrum suggested that **1** is a coumarin. Additional evidence for the presence of this carbon skeleton came from the 1H NMR spectrum. The signals corresponding to C-3 and C-4 protons appeared as an AB system at $\delta 6.30$ and 7.63 (J

= 9.8 Hz). Two singlets at $\delta 7.00$ and 7.38 (1H each) were attributable to C-8 and C-5 protons, respectively. The 1H NMR spectrum also showed the presence of two acetyl groups at $\delta 2.07$ (3H \times 2, s) and one methoxyl group at $\delta 3.70$ (3H, s).

On acetylation with acetic anhydride–pyridine, **1** gave a tetra-acetate (**2**), mp 168–169°, $C_{24}H_{26}O_{13}$. The physical and spectral data of **2** [$\nu_{max}^{nujol} cm^{-1}$: 1770–1730, 1620, 1570, 1505, 920, 890 and 825; $\delta 2.10$ (3H \times 3, s), 2.17 (3H, s), 3.95 (3H, s), 7.20, 7.40 (1H each, s), 6.38 and 7.95 (1H each, d, $J = 10$ Hz)] were identical with those of scopolin acetate [3].

Therefore, two acetyl groups in **1** are located in the glucosyl moiety, and their positions were determined by detailed analysis of the 1H NMR spectrum of **1** with aid of decoupling procedures. A doublet at $\delta 5.43$ (1H, $J = 8.5$ Hz) was assigned to an anomeric proton, which was coupled with a C'-2 proton at $\delta 5.68$ (1H, dd, $J = 8.5$ and 8.5 Hz). On irradiation at $\delta 5.68$, a double doublet at $\delta 4.23$ (1H, dd, $J = 8$ and 8.5 Hz) due to a C'-3 proton collapsed